Note

Amylose percentage and distribution of unit chain-length of maize starches having a specific genetic background*†

YOSHIKO IKAWA, DAVID V. GLOVER[‡], YOSHIMI SUGIMOTO, AND HIDETSUGU FUWA

Department of Food and Nutrition, Osaka City University, Sugimotocho, Sumiyoshi-ku, Osaka (Japan)

(Received June 30th, 1977; accepted for publication in revised form, September 2nd, 1977)

In this study, starch granules prepared from kernels of the single mutants, amylose-extender (ae), waxy (wx), and opaque-2 (o_2), and the double-mutant combinations ae o_2 and wx o_2 from four maize (Zea mays L.) inbred lines, Oh43, B37, C103, and W64A were fractionated by gel filtration after debranching by isoamylase. The contents of each fraction and distribution of the linear α -D-(1 \rightarrow 4)-linked unit-chains were investigated. No noteworthy differences were observed in the aforementioned criteria between single mutants and their respective double-mutant combinations with o_2 , and among four inbred lines. It was confirmed that starch granules of the ae mutants have intermediate fraction(s) different from typical amylose and amylopectin.

Several of the endosperm mutants of maize (Zea mays L.) display differences in pasting temperature, BEPT**, viscosity, gel-stability, starch-granule digestibility, and the production of amylose, amylopectin, water-soluble polysaccharides, and sugars (see Fuwa et al.¹ for references). Greenwood and his co-workers² found that iodine-binding curves for starches of amylose-extender (ae) mutants were different from that of normal maize-starch, and amylose contents estimated by potentiometric iodine-titration were higher (by about 15%) than those of complexes with 1-butanol. Furthermore, they² separated from the ae starches an intermediate fraction different from typical amylose and amylopectin. It is remarkable that the ae starches had large portions of the intermediate fraction, whereas normal maize-starches did not. Recently, Boyer et al.³ and Yamada et al.⁴ reported that maize starches of the amylose-extender waxy (ae wx) double-mutant consists merely of an atypical amylopectin. They suggested the abnormal amylopectin was similar to the intermediate fraction

^{*}Dedicated to Professor Dexter French on the occasion of his 60th birthday. The main part of this paper was presented at the Annual Meeting of the Japanese Society of Starch Science, Tokyo, August 25, 1976, at which Professor French delivered a special lecture.

[†]Journal Paper no. 6754, Purdue Agricultural Experiment Station, West Lafayette, IN 47907 (U.S.A.)
†Department of Agronomy, Purdue University, West Lafayette, IN 47907, U.S.A.

^{**}BEPT denotes Birefringence Endopoint Temperature.

212 NOTE

of the ae starches. Thus, in investigating components of maize starches, it is necessary to account not only for typical amylose and amylopectin, but also for these intermediate fraction(s).

Our investigations on susceptibility of various starch granules to amylases required fuller knowledge about starch components. To help elucidate the details, we prepared starch granules from kernels of several single mutants and their double-mutant combinations having o_2 of four, inbred lines of maize, and measured their content of amylose and the distribution of linear, α -D-(1 \rightarrow 4)-linked, unit-chains by the use of gel filtration after debranching by isoamylase.

RESULTS AND DISCUSSION

Figs. 1A and 1B show typical elution patterns on a Sephadex G-75 column of maize starch after debranching by isoamylase. A peak appeared near the void volume [fraction I (Fr. I)], a peak or shoulder near the average chain-length (c.l.) of 50 [fraction II (Fr. II)], and a peak at c.l. 20 [fraction III (Fr. III)]. Fr. I comprised material of higher molecular weight, regarded as amylose in the original granules. Frs. II and III constituted the linear unit-chains of amylopectin, because they displayed peaks at about c.l. 50 and 20, respectively.

Table I shows that there are no significant differences apparent in the contents of each of Frs. I, II, and III in the percentage and chain length of each of the apices from Frs. II and III, whether the high-amylose maize starches are debranched by isoamylase or by a combination of isoamylase and pullulanase. In addition, there were no differences between their elution patterns, and thus we used isoamylase for debranching all other starches.

Fig. 1D shows that the commercial, waxy-maize starch contains a trace of Fr. I. However, in the wx and wx o_2 starches of Oh43 and B37, there is no Fr. I (Table II). The elution patterns of wx and wx o_2 starches of Oh43 and B37 resemble that of the commercial wx starches very closely, except Fr. I. As these starches of the wx mutants consist essentially of amylopectin, the contents and elution patterns of Frs. II and III characterize typical amylopectin. As compared with the elution-

TABLE I

PERCENTAGE OF FRACTIONS I, II, AND III, AND AVERAGE CHAIN-LENGTH (C.L.) OF HIGH-AMYLOSE MAIZE STARCH, DEBRANCHED BY ISOAMYLASE OR ISOAMYLASE PLUS PULLULANASE

Debranching enzymes	Fr. I (%)	Fr. II (%)	Fr. III (%)	Frs. II+III (%)		Peak of Fr. III (c.l.)
Isoamylase Isoamylase plus	44.0	36.8	19.2	56.0	52	18
pullulanase	42.8	36.0	21.2	57.2	45	18

213

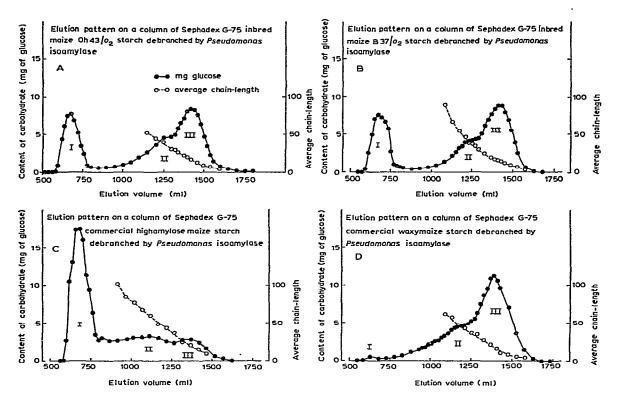


Fig. 1. Elution patterns on a column of Sephadex G-75 of four maize starches debranched by iso-amylase: (A) opaque-2 (O₂) maize starch of Oh43, (B) O₂ maize starch of B37, (C) commercial high-amylose VII maize starch, (D) commercial waxy maize starch (———) content of carbohydrate determined by the phenol-sulfuric acid method⁵ and expressed as mg of glucose; average length of unit chain (—O—) calculated from the carbohydrate content and the number of reducing ends, as determined by the chromotropic acid method^{6,7}).

patterns of the o_2 and wx or wx o_2 (see Fig. 1), Frs. II and III displayed similar elution patterns. Apparently, the o_2 starches contained typical amylopectin.

Fig. 1C shows that the high-amylose VII starches had higher contents of carbohydrate between Fr. I and Fr. II than those of the o_2 mutants. Starches of the ae and ae o_2 mutants of the Oh43, B37, C103, and W64A maize inbred-strains were similar in their elution profiles to that of the high-amylose VII starch. This indicates that the high-amylose maize starches contained intermediate fraction(s) different from typical amylose and amylopectin. The clution patterns of Frs. II and III of the high-amylose maize starches were also different from those of the o_2 and wx mutants. Apparently, the high-amylose maize starches contained an anomalous amylopectin and/or an amylose of short chain-length.

Table II shows the percentage contents of Frs. I, II, and III, and the chain lengths of apices in Frs. II and III. The contents of Fr. I were about 28% of the o_2 , 0% of the wx and wx o_2 , and 1% of the commercial waxy-maize starch, respectively,

214 NOTE

TABLE II

PERCENTAGE OF FRACTIONS I, II, AND III, AND AVERAGE CHAIN LENGTH (C.L.) OF APICES OF PEAKS II AND III

Inbred line	Genotype	Fr. I	Fr. II	Fr. III	Frs. II+I	II Peak of	Peak of
		(%)	(%)	(%)	(%)	$Fr.II(\overline{c.l.})$	Fr. III (c.l.)
	02	28.3	26.4	45.3	71.7	40	17
	wx	0	32.1	67.9	100.0	45	17
OH43	wx 02	0	36.5	63.5	100.0	46	18
	ae	39.6	40.7	19.7	60.4	53	18
	ae 02	39.0	36.8	24.2	61.0	55	17
	02	28.2	21.7	50.1	71.8	40	17
	wx	0	33.9	66.1	100.0	44	17
B37	WX 02	0	34.9	65.4	100.0	48	18
	ae	38.0	34.7	27.3	62.0	45	18
	ae 02	37.0	36.0	27.0	63.0	45	19
C103	ae	34.6	38.7	26.7	65.4	40	17
	ae o_2	35.3	33.2	31.5	64.7	43	17
W64A	ae	36.7	36.1	27.2	63.3	44	18
	ae 02	39.3	34.0	26.8	60.8	43	18
Commercial	HA*	55.0	31.5	13.5	45.0	60	20
	waxy	1.1	36.3	63.7	98.9	48	19

^{*}Different lots of commercial, high-amylose maize starch from those used in Table I.

and agreed with the amylose percentage as measured by potentiometric iodine-titration⁸. In the high-amylose maize starches, the contents of Fr. I were 55% of the high-amylose VII, and 35-40% of the ae and ae o_2 starches in the four inbred lines. These values were 10-15% lower than the amylose percentage measured by the other methods^{8,9}. This fact, as pointed out by Greenwood and his co-workers² and by Boyer et al.³, related to the aforementioned intermediate fraction(s), and this is a striking characteristic of the high-amylose maize starches. However, the elucidation of the relationship between the results obtained and the digestibility of starch granules needs further study. There were no significant differences in the chain length of apices of Frs. II and III among the four inbred lines of maize, and between starches of the double-mutant combinations having o_2 and those of their respective, nonopaque single mutants.

EXPERIMENTAL

Materials. — Samples of starch granules were prepared from mature maize (Zea mays L.) kernels of the ae and ae o_2 mutants of inbreds Oh43, B37, C103, and W64A, and the o_2 , wx and wx o_2 mutants of Oh43 and B37. Starches were prepared according to Schoch's method¹⁰. As controls, commercial high-amylose VII maize

NOTE 215

starch (Shikishima Denpun Co., Japan) and waxy-maize starch (Clifford Lobe Co., Ltd., Australia) were used.

Methods. — Starches were debranched by the method of Mercier and Kainuma11, and debranched starches were fractionated according to the method of Akai et al.6. Starch granules (a definite wet-weight between 450 and 455 mg) were dissolved in 10 ml of dimethyl sulfoxide, incubated for 1 h at 40°, and 20 ml of 0.01M acetate buffer, pH 3.5, and 8850 U* of crystalline Pseudomonas isoamylase (Hayashibara Biochemical Laboratories, Inc., Okayama, Japan) were added. The mixture was incubated for 24 h at 40° for debranching. When both isoamylase and pullulanase (Hayashibara Biochemical Laboratories, Inc., Okayama, Japan) were used for debranching, further incubation under similar conditions was performed with 10 ml of 0.5m acetate buffer, pH 5.5, and 40 IU[†] of crystalline pullulanase. Five-fold volumes of ethanol were added to the mixture and starches were precipitated. The precipitates were washed twice with ethanol and then twice with ether, and dried in vacuo. The dried samples were suspended in 5 ml of water, and gelatinized with 1 ml of 6M sodium hydroxide under nitrogen. After 1 h, ungelatinized materials were centrifuged off (1,200g 10 min). The supernatant was diluted to 10 ml with water and crystalline sodium chloride added to a concentration of 5% (w/v). These samples (10 ml each) were charged onto a column (5.0 × 85 cm) of Sephadex G-75 and eluted with 0.05м sodium hydroxide containing 5% sodium chloride. Fractions (10 ml) were collected, and neutralized with 0.5M hydrochloric acid. The amount of carbohydrate in each tube was measured by the phenol-sulfuric acid method⁵. Reducing end-groups were determined colorimetrically by the chromotropic acid method, according to Akai et al.6,7. The average chain length (c.l.) was calculated from the amount of carbohydrate and number of reducing ends. The contents of each fraction (Frs. I, II, and III, see Results and Discussion) were calculated from the elution areas.

ACKNOWLEDGMENTS

The authors express their gratitude to Dr. Kainuma of National Food Research Institute, Tokyo, Japan, for a preprint of his unpublished work. This investigation was supported, in part, by a grant from the Ministry of Education of Japan and by the Agency for International Development under contract, "Inheritance and Improvement of Protein Quality and Content in Maize".

REFERENCES

- 1 H. FUWA, M. NAKAJIMA, A. HAMADA, AND D. V. GLOVER, Cereal Chem., 54 (1977) 230-237.
- 2 W. Banks, C. T. Greenwood, and D. D. Muir Chorleywood, Die Stärke, 26 (1974) 289-328.
- 3 C. D. BOYER, D. L. GARWOOD, AND J. C. SHANNON, Die Stärke, 28 (1976) 405-410.

^{*}One unit is expressed according to ref. 12.

^{*}One unit is the amount of pullulanase that releases 1 μ mol of maltotriose from pullulan in 1 min at pH 5.0 and 30°.

216

- 4 T. YAMADA AND M. TAKI, Die Stärke, 28 (1976) 374-377.
- 5 M. Dubois, K. A. Gilles, J. K. Hamilton, P. A. Rebers, and F. Smith, *Anal. Chem.*, 28 (1956) 350-356.
- 6 H. AKAI, K. YOKOBAYASHI, A. MISAKI, AND T. HARADA, Biochim. Biophys. Acta, 237 (1971) 422-429.
- 7 D. J. HANAHAN AND J. N. OLLEY, J. Biol. Chem., 231 (1958) 813-828.
- 8 H. Fuwa, Denpun Kagaku, 20 (1973) 120-130.
- 9 G. K. ADKINS AND C. T. GREENWOOD, Carbohydr. Res., 3 (1966) 152-156.
- 10 T. J. Schoch, Methods Enzymol. 3 (1954) 6-7.
- 11 C. MERCIER AND K. KAINUMA, Die Stärke, 27 (1975) 289-292.
- 12 K. Yokobayishi, A. Misaki, and T. Harada, Biochim. Biophys. Acta, 212 (1970) 458-469.